New cross-talk measure of near-infrared spectroscopy and its application to wavelength combination optimization

Shinji Umeyama Toru Yamada

National Institute of Advanced Industrial Science and Technology Neuroscience Research Institute 1-1-1 Umezono Central 2 Tsukuba-shi, Ibaraki 305-8568 Japan E-mail: s.umeyama@aist.go.jp Abstract. In near-infrared spectroscopy, concentration changes in oxygenated and deoxygenated hemoglobin are calculated from the changes in the attenuation of the measurement light. This is done by solving a linear equation based on the modified Lambert-Beer law. To solve this equation, we need to know the partial optical pathlengths in the activated region in the brain. Because they are difficult to know, a wavelength-independent constant or a wavelength-dependent total optical pathlength has been substituted for these values in actual measurements. This kind of substitution inevitably produces errors, called cross-talk, when calculating concentration changes. In this paper, we propose a new cross-talk measure for dual and triple wavelength measurements, and analyze it over various wavelength combinations. The results indicate that constant substitution is not inferior to total pathlength substitution in dual wavelength measurements, and that total path-length substitution is very effective for triple wavelength measurements. © 2009 Society of Photo-Optical Instrumentation Engineers. [DOI: 10.1117/1.3147402]

Keywords: near-infrared spectroscopy; cross-talk; Monte Carlo simulation. Paper 08388R received Nov. 20, 2008; revised manuscript received Mar. 2, 2009;

accepted for publication Mar. 25, 2009; published online Jun. 8, 2009.

1 Introduction

Near-infrared spectroscopy (NIRS) is an effective tool for non-invasive investigation of cerebral oxygenation and hemodynamics during functional brain activation.^{1–3} It has several advantages over other functional measurement methods such as functional magnetic resonance imaging, positron emission tomography, and electroencephalography. These advantages include good temporal resolution, measurement of both oxygenated (HbO) and deoxygenated (HbR) hemoglobin, portability, and low equipment cost.⁴

In NIRS, a simple calculation based on measurements of the change in the attenuation of light as it propagates through the head of a subject can provide the concentration change of HbO and HbR in the brain. The relationship between attenuation and concentration change is modeled by a linear equation based on the modified Lambert–Beer law. The coefficients of the linear equation include the molar absorption coefficients of HbO and HbR,⁵ and the optical pathlengths of the measurement light in the activated region. If the oxy- and deoxy-hemoglobin concentrations change homogeneously throughout the entire tissue volume, a total optical pathlength can be used as the coefficient. However, if these concentrations change locally in the cortex, a partial optical pathlength of the cortical tissue should be used to obtain an accurate estimation.^{6–8} The partial optical pathlengths of the cortical

tissue are difficult to know because there is no experimental method of directly obtaining these values. Thus, historically, a wavelength-independent constant (unity) or a wavelength-dependent total optical pathlength has been substituted for these values. This kind of substitution, however, inevitably provides a source of error when calculating concentration changes. These errors are referred to as *cross-talk* because a change in one of the chromophores may mimic a change in another chromophore.^{6,7}

If we adopt a constant or a total pathlength as a substitution for the partial pathlength, cross-talk is inevitable. Thus, it is important to select a good combination of measurement light wavelengths to minimize the effect. Uludag et al.⁶ proposed one measure of cross-talk, which led to other studies reporting on good wavelength combinations.^{9,10} However, Uludag's measure may be inadequate in some situations because it gives a zero value (no cross-talk) when an estimation error really exists.

In this paper, we propose a new cross-talk measure, which provides a unified treatment of both HbO and HbR crosstalks. This measure is guaranteed to be zero if and only if there is no cross-talk. The proposed values and Uludag's values of dual and triple wavelength measurements for various wavelength combinations were calculated and compared in two cases—a wavelength-independent constant substitution and a wavelength-dependent total optical pathlength substitution. An appropriate wavelength combination, which minimizes cross-talk error, was given. The cross-talk calculation

Address all correspondence to Shinji Umeyama, Neuroscience Research Institute, National Institute of Advanced Indusrial Science and Technology (AIST) Umezono 1-1-1-Central 2, Tsukuba, Ibaraki 305-8568, Japan; Tel: 81-29-861-5837; Fax: 81-29-861-5841; E-mail: s.umeyama@aist.go.jp

^{1083-3668/2009/14(3)/034017/8/\$25.00 © 2009} SPIE

was performed based on the estimated total and partial optical pathlength obtained by Monte Carlo simulation on a layered model of an adult head.

2 Theory

2.1 NIRS Method

When using near-infrared light in a uniformly turbid medium, the temporal attenuation change ΔA resulting from a small homogeneous change in the absorption coefficient $\Delta \mu_{\alpha}$ can be represented by the modified Lambert–Beer law with the assumption that measurement noise is absent

$$\Delta A(\lambda) = l(\lambda) \Delta \mu_a(\lambda), \qquad (1)$$

where $l(\lambda)$ is the optical pathlength of the activated region at wavelength λ , and $\Delta \mu_a$ is given as

$$\Delta \mu_{a}(\lambda) = \varepsilon_{\text{HbO}}(\lambda) \Delta \text{HbO} + \varepsilon_{\text{HbR}}(\lambda) \Delta \text{HbR}.$$
 (2)

In Eq. (2), Δ HbO and Δ HbR represent oxygenated and deoxygenated hemoglobin concentration changes, whereas $\epsilon_{\text{HbO}}(\lambda)$ and $\epsilon_{\text{HbR}}(\lambda)$ denote their molar absorption coefficients.

Estimating the concentration changes in two kinds of chromophores (Δ HbO and Δ HbR) requires at least two light sources of different wavelengths ($\lambda_1, \lambda_2, ..., \lambda_m, m \ge 2$). Dual (m=2) and triple (m=3) wavelength measurements are popular in NIRS. We use vectors to represent the temporal attenuation change and hemoglobin concentration changes as

$$\boldsymbol{a} = [\Delta A(\lambda_1), \Delta A(\lambda_2), \dots, \Delta A(\lambda_m)]^T,$$
(3)

$$\boldsymbol{x} = (\Delta \text{HbO}, \Delta \text{HbR})^T.$$
(4)

Equations (1) and (2) are summarized as follows:

$$a = LEx, \tag{5}$$

where

$$L = \operatorname{diag}[l(\lambda_1), l(\lambda_2), \dots, l(\lambda_m)], \qquad (6)$$

$$E = \begin{pmatrix} \varepsilon_{\rm HbO}(\lambda_1) & \varepsilon_{\rm HbR}(\lambda_1) \\ \varepsilon_{\rm HbO}(\lambda_2) & \varepsilon_{\rm HbR}(\lambda_2) \\ \dots & \dots \\ \varepsilon_{\rm HbO}(\lambda_m) & \varepsilon_{\rm HbR}(\lambda_m) \end{pmatrix}.$$
 (7)

If hemoglobin concentrations change locally in the cortex during brain activation, the temporal attenuation change a should be given as follows.^{11,12}

$$\boldsymbol{a} = L_{\text{cortex}} \boldsymbol{E} \boldsymbol{x},\tag{8}$$

where

1

$$L_{\text{cortex}} = \text{diag}[l_{\text{cortex}}(\lambda_1), l_{\text{cortex}}(\lambda_2), \dots, l_{\text{cortex}}(\lambda_m)], \quad (9)$$

and l_{cortex} is a partial optical pathlength of the cortical tissue.

If the partial optical pathlength within the cortex is known in advance, we can obtain accurate values of hemoglobin concentration changes by applying the inverse or pseudo inverse matrix of $L_{\text{cortex}}E$ to the temporal attenuation change \boldsymbol{a} . However, it is difficult to know this pathlength in advance since the partial optical pathlength cannot be measured experimentally. In practice, we approximate L_{cortex} using a known matrix $L_s = \text{diag}(l_1, l_2, \dots, l_m)$. Thus, the estimated hemoglobin concentration change $\hat{\boldsymbol{x}}$ is given as

$$\hat{\boldsymbol{x}} = \boldsymbol{E}^+ \boldsymbol{L}_{\rm s}^{-1} \boldsymbol{L}_{\rm cortex} \boldsymbol{E} \boldsymbol{x} \tag{10}$$

$$=T(L_{\rm s})\boldsymbol{x}.\tag{11}$$

We refer to the matrix, $T(L_s)$, as a *reproduction matrix* of hemoglobin concentration change. A wavelength-independent constant matrix I (a unit matrix) or a wavelength-dependent total path-length matrix L_{total} have historically been used as L_s . We represent

$$T_{\rm c} = T(I) = E^+ L_{\rm cortex} E, \qquad (12)$$

$$T_{\rm t} = T(L_{\rm total}) = E^+ L_{\rm total}^{-1} L_{\rm cortex} E, \qquad (13)$$

in this paper.

If we set $L_s = L_{cortex}$, the reproduction matrix *T* becomes a unit matrix. Thus, its reproduction is complete and the hemoglobin concentration changes obtained is equal to their true values. However, if we set $L_s = I$ or $L_s = L_{total}$, *T* is no longer a unit matrix. Therefore, it leads to cross-talk errors in the derived concentration changes.

2.2 Uludag's Cross-Talk Measure

Uludag et al.⁶ proposed a measure to evaluate cross-talk caused by a deformed reproduction matrix. They defined the cross-talk $C_{A\rightarrow B}$ from chromophore A to chromophore B as the ratio of the determined concentration change of chromophore B and A, where a change was introduced in A and not in B. Thus, Uludag's cross-talk measure represented by a reproduction matrix *T* are given as follows:

$$C_{\text{HbO}\to\text{HbR}}(T) = \frac{T_{21}}{T_{11}},$$
 (14)

$$C_{\text{HbR}\to\text{HbO}}(T) = \frac{T_{12}}{T_{22}},$$
 (15)

where

$$T = \begin{pmatrix} T_{11} & T_{12} \\ T_{21} & T_{22} \end{pmatrix}.$$
 (16)

2.3 New Cross-Talk Measure

Here we propose a new cross-talk measure. Because we do not know the real relationship between the concentration changes of HbO and HbR, we use here a very simple assumption that hemoglobin concentration changes reside on a unit circle. We evaluate the mean squared error of the estimated hemoglobin concentration changes induced by a reproduction matrix T.

Let **x** be a point on a unit circle $(0 \le \theta \le 2\pi)$,

$$\boldsymbol{x}(\theta) = \begin{pmatrix} \cos \theta \\ \sin \theta \end{pmatrix}.$$
 (17)

Then the mean squared error J(T) of the reproduced hemoglobin concentration change $\hat{x} = Tx$ is defined as

$$J(T) = \min_{w} \frac{1}{2\pi} \int_{0}^{2\pi} \|w \hat{\mathbf{x}} - \mathbf{x}\|^{2} \mathrm{d}\theta.$$
(18)

At this point, we introduced a scaling factor, w, because the estimated hemoglobin concentration change has an ambiguity in its scaling.

Let the integral term in Eq. (18) be g(w),

$$g(w) = \frac{1}{2\pi} \int_0^{2\pi} \|wT\mathbf{x}(\theta) - \mathbf{x}(\theta)\|^2 \mathrm{d}\theta$$
(19)

$$=\operatorname{tr}\left\{ (wT-I)^{T}(wT-I) \cdot \frac{1}{2\pi} \int_{0}^{2\pi} \boldsymbol{x}(\theta) \boldsymbol{x}(\theta)^{T} \mathrm{d}\theta \right\}$$
(20)

$$= \frac{1}{2} [w^2 ||T||^2 - 2w \operatorname{tr}(T) + 2], \qquad (21)$$

where we used

$$\frac{1}{2\pi} \int_0^{2\pi} \boldsymbol{x}(\theta) \boldsymbol{x}(\theta)^T \mathrm{d}\theta = \int_0^{2\pi} \begin{pmatrix} \cos^2 \theta & \sin \theta \cos \theta \\ \sin \theta \cos \theta & \sin^2 \theta \end{pmatrix} \mathrm{d}\theta$$
(22)

$$=\frac{1}{2}I.$$
 (23)

Then, the minimum of g(w) is achieved when

$$w^* = \frac{\operatorname{tr}(T)}{\|T\|^2}.$$
 (24)

Substituting this value in Eq. (21), we have

$$g(w^*) = 1 - \frac{\operatorname{tr}(T)^2}{2||T||^2}.$$
 (25)

Thus,

$$J(T) = 1 - \frac{\operatorname{tr}(T)^2}{2||T||^2}.$$
 (26)

We propose J(T) as a new cross-talk measure. By the given definition, J(T)=0 if and only if T=cI. J(T) is also scale invariant [i.e., J(ct)=J(T)].

2.4 Geometric Interpretation of Cross-Talk Measures

Here, we give geometric interpretations of the cross-talk measure along with that provided by Uludag. If the true hemoglobin concentration change x resides on a unit circle, then the reproduced hemoglobin concentration change $[x' = w^*Tx(\theta)]$ is on an ellipsoid. Figure 1(a) shows these two distributions. The blue line shows the distribution of x, and the green line

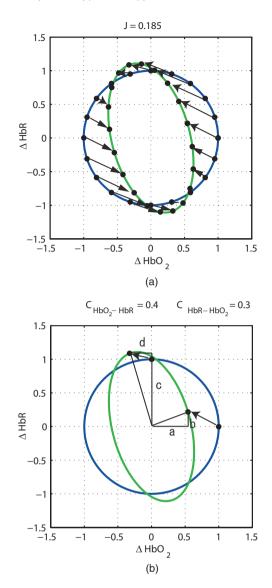


Fig. 1 Geometric interpretation of cross-talk measures. The blue circle represents the original concentration change, and the green ellipsoid represents the reproduced concentration change: (a) The proposed measure and (b) Uludag's measure.

shows the corresponding distribution of x'. Here, we used the following reproduction matrix T, as an example,

$$T = \begin{pmatrix} 1.0 & -0.6\\ 0.4 & 2.0 \end{pmatrix}.$$
 (27)

The scale value $w^*=0.5435$ in this case. The arrows in Fig. 1(a) represent the reproduction from x to x'. Thus, our crosstalk measure J(T) corresponds to the mean squared sum of these arrow lengths. Conversely, Fig. 1(b) shows a geometric interpretation of the Uludag measure. Blue and green lines indicate the same distributions as in Fig. 1(a). Two points on a unit circle, $x_1=(1,0)^T$ and $x_2=(0,1)^T$, move to the reproduced points along the arrows shown in Fig. 1. Thus, $C_{\text{HbO}\rightarrow\text{HbR}}=b/a$ and $C_{\text{HbR}\rightarrow\text{HbO}}=d/c$.

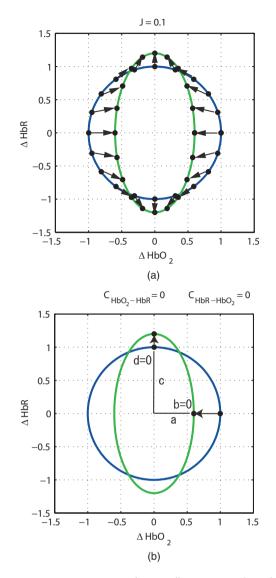


Fig. 2 Geometric interpretation of cross-talk measures, where the Uludag's measure results in zero despite the existence of reproduction error: (a) The proposed measure gives an appropriate cross-talk value and (b) Uludag's measure indicates no cross-talk.

The Uludag cross-talk measures were determined by deformations at only two points; therefore, it fails to give an appropriate deformation measure in certain situations. For example, see Fig. 2(b), where

$$T = \begin{pmatrix} 1.0 & 0\\ 0 & 2.0 \end{pmatrix} \tag{28}$$

and $w^*=0.6$. In this case, the Uludag measures are $C_{\text{HbO}\rightarrow\text{HbR}}=0$ and $C_{\text{HbR}\rightarrow\text{HbO}}=0$ even though *T* obviously gives deformation. This implies that *T* is not necessarily equal to *cI* (no deformation) even if $C_{\text{HbO}\rightarrow\text{HbR}}=0$ and $C_{\text{HbR}\rightarrow\text{HbO}}=0$. On the other hand, our measure gives an appropriate evaluation of deformation even in this case [see Fig. 2(a)].

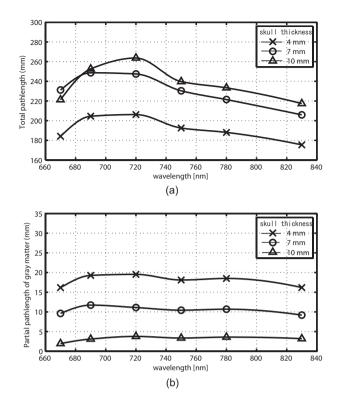


Fig. 3 Wavelength dependence of (a) the total optical pathlength and (b) the partial optical pathlength in the cortex. The skull thickness was assumed to be 4, 7, and 10 mm, and the source-detector spacing was set to 30 mm. pathlengths at six wavelengths were predicted by Monte Carlo simulation and interpolated to obtain the curves.

3 Determining Wavelength-Dependent Optical Pathlength by Monte Carlo Simulation

A cross-talk value depends on the wavelength combination of source lights because both molar absorption coefficients and optical pathlengths depend on the wavelength. To study this relation, we performed a Monte Carlo simulation and predicted the wavelength-dependent total and partial optical pathlengths. We used tMCimg, the simulation program provided to public by the Photon Migration Imaging Laboratory at Massachusetts General Hospital. A a five-layer model of an adult head consisting of scalp, skull, cerebrospinal fluid (CSF), gray matter, and white matter was used. The size of simulated tissues was $100 \times 100 \times 50$ mm, and the thicknesses of the scalp, CSF, and gray matter layers were 3, 2, and 4 mm, respectively. Because the skull thickness has a great influence on the pathlength, three simulation trials based on different thicknesses (4, 7, and 10 mm) of the skull layer were performed. The source-detector separation was set 30 mm.

The simulation was performed for six wavelengths (670, 690, 720, 750, 780, and 830 nm), and the total and partial optical pathlengths at each wavelength were calculated. The total and partial optical pathlengths for other wavelengths were determined by interpolating these six values. The wavelength-dependent optical properties (absorption coefficient μ_a and transport scattering coefficient μ'_s) at six wavelengths are shown in Table 1. These data are drawn from a recent report.¹⁰

Figure 3 shows the total pathlength [Fig. 3(a)] and the

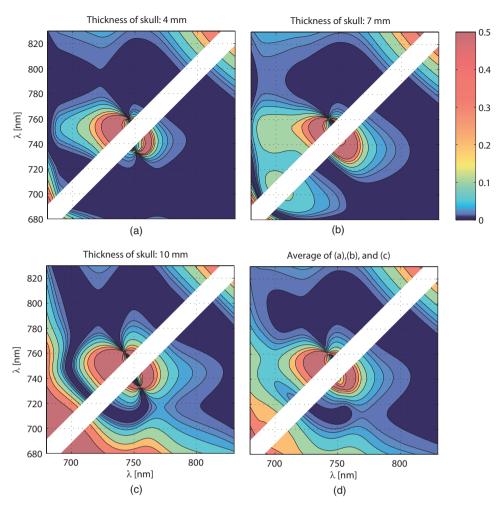


Fig. 4 Proposed cross-talk by dual wavelength measurement. Combinations of two wavelengths between 680 and 830 nm are shown. (a, b, c) correspond to skull thicknesses of 4, 7, and 10 mm, respectively. The upper triangle represents cross-talk using a constant substitution $[J(T_c)]$. The lower triangle represents cross-talk using a total pathlength substitution $[J(T_t)]$. (d) shows the average of (a), (b), and (c).

partial pathlength in the cortex [Fig. 3(b)] obtained by our simulation. These values were used for our calculations of cross-talk.

4 Results and Discussion

Figures 4(a)-4(c) shows the proposed cross-talk measure values [Eq. (26)] calculated from the Monte Carlo simulation result. Dual wavelength measurement was assumed. Figures 4(a)-4(c) correspond to skull thicknesses assumed to be 4, 7, and 10 mm, respectively. The upper triangle of each figure shows the cross-talk $J(T_c)$, where a unit matrix I was used as $L_{\rm s}$. The lower triangle gives the $J(T_{\rm t})$, where the total pathlength matrix L_{total} was used. The cross-talk values were generated for all combinations of the two wavelengths between 680 and 830 nm, except when two wavelengths were too close to each other. The colors denote the cross-talk values, ranging from 0 (blue) to 0.5 (red). In this way, good wavelength combinations having a small cross-talk are represented by dark blue. Fig. 4(d) shows the average for the subfigures. The dark blue area of Fig. 4(d) indicates good wavelength combinations-those that provide little cross-talk for a wide range of skull thickness.

Figures 4(a) and 4(b) indicate that the cross-talks are similar for skull thickness of 4 and 7 mm. On the other hand, a skull thickness of 10 mm [Fig. 4(c)] showed different cross-talk. This is likely because changes in partial optical pathlength with respect to the wavelength are large when the skull thickness is 10 mm.

The dark blue area in Fig. 4(d) indicates good wavelength combinations, producing little cross-talk for a wide range of skull thicknesses. These combinations are different for $J(T_c)$ (upper triangle) and $J(T_t)$ (lower triangle). Interestingly, however, the area sizes are similar. Therefore, a simple substitution of a constant is not an inferior method to a total pathlength substitution if the wavelength combination is carefully selected.

The cross-talk for triple wavelength measurement is shown in Figs. 5(a)-5(c). In our analysis, one wavelength among the three source lights was fixed at 830 nm. The cross-talk for all combinations of the two remaining wavelengths between 680 and 820 nm, except when the two wavelengths were too close to each other, are given. The parameters used to draw the figure and the meaning of each subfigure are the same as for Fig. 4. Figure 5(d) shows the average for the subfigures.

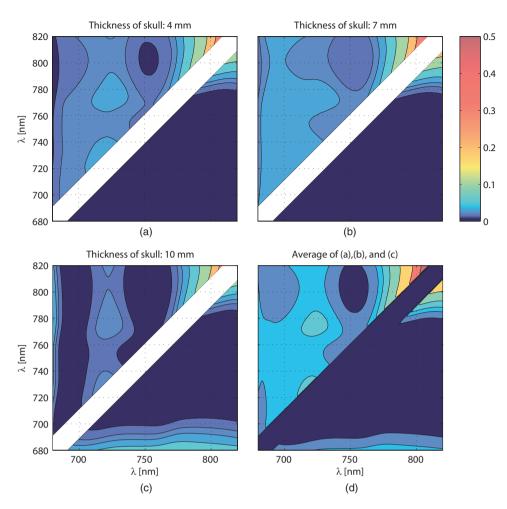


Fig. 5 Proposed cross-talk by triple wavelength measurement. One wavelength was fixed 830 nm. Combinations of other two wavelengths between 680 and 820 nm were tested and are shown. (a, b, c) correspond to skull thickness of 4, 7, and 10 mm, respectively. The upper triangle represents cross-talk using a constant substitution $[J(T_c)]$. The lower triangle represents cross-talk using a total pathlength substitution $[J(T_t)]$. (d) shows the average of (a), (b), and (c).

Figure 5(d) shows that triple wavelength measurement provides good cross-talk performance for many wavelength combinations if we use a total path-length substitution. On the other hand, triple wavelength measurement using a constant substitution gives a very good cross-talk performance for a small number of combinations. Thus, we need to carefully select the wavelength combination if we use a constant substitution.

The absolute values of Uludag's cross-talk measure, $C_{\text{HbO}\rightarrow\text{HbR}}$, are shown in Fig. 6(a) for dual wavelength measurement and in Fig. 6(b) for triple wavelength measurement. Figures 6(a) and 6(b) show the averaged cross-talk values when skull thickness values of 4, 7, and 10 mm were assumed. Because the scaling of Uludag's measure is different from the proposed one, its value was scaled so that both the proposed and Uludag's measure give an identical cross-talk value at the wavelength combination of 690 and 830 nm in Fig. 6(a) and 780, 805, and 830 nm in Fig. 6(b), respectively.

In dual wavelength measurement, the cross-talk of the proposed measure [Fig. 4(d)] and Uludag's [Fig. 6(a)] are basically similar. However, when two wavelengths are small (lower left corner of each figure), Uludag's measure gives

very small cross-talk values compared to the proposed method. A weak tendency to do this is also found in triple wavelength measurement.

5 Conclusion

Simulation results show the following:

1. Triple wavelength measurement with a total path-length substitution is the most robust method of selecting the wavelength combination because many wavelength combinations show little cross-talk.

2. Constant and total path-length substitutions in dual wavelength measurement give similar performance because the dark blue area in Fig. 4(d) (wavelength combinations producing little cross-talk) is roughly symmetric with respect to the diagonal line of the figure. Thus, a constant substitution may be more favorable because it is applicable without knowing the total pathlength.

3. Uludag's measure usually gives similar cross-talk to the proposed measure, except the case where the two wavelengths are small concurrently in dual wavelength measurement.

Absorption coefficient μ_{α} (mm ⁻¹)						
Wavelength (nm)	670	690	720	750	780	830
Scalp	0.031	0.028	0.022	0.022	0.020	0.019
Skull	0.027	0.017	0.013	0.016	0.016	0.017
CSF	0.0044	0.0029	0.0030	0.0045	0.0044	0.0056
Gray matter	0.048	0.039	0.036	0.038	0.036	0.041
White matter	0.024	0.018	0.014	0.016	0.016	0.018
		Transport scatte	ring coefficient $\mu_{ m s}'$ (mm ⁻¹)		
Wavelength (nm)	670	690	720	750	780	830
Scalp	2.53	2.38	2.24	2.11	2.00	1.84
Skull	2.33	2.13	1.92	1.79	1.66	1.47
CSF	0.35	0.32	0.29	0.27	0.25	0.22
Gray matter	2.69	2.57	2.46	2.39	2.31	2.10
White matter	10.49	10.03	9.78	9.62	9.25	8.82

 Table 1
 Wavelength-dependent optical properties (absorption coefficient and transport scattering coefficient) and the thickness of each layer of our head model. These values were drawn from Ref. 10.

The proposed measure can be easily extended to the crosstalk problem, including more than three chromophores. Uludag's measure is also extendable. However, if three chromophores (for example, A, B, and C) are considered, six cross-talk measures are defined by Uludag's approach: $C_{A\rightarrow B}, C_{B\rightarrow A}, C_{A\rightarrow C}, C_{C\rightarrow A}, C_{B\rightarrow C}$, and $C_{C\rightarrow B}$. Thus, we may need an another criterion to integrate these measures into one.

Measurement noise is another important factor in selecting a wavelength combination because it significantly influences the measurement accuracy. Several studies concerning this issue have been performed,^{13,9} and they pointed out that two wavelengths that are separated from each other basically give good performance in dual wavelength measurement. Thus, we should select a wavelength combination that is in the small cross-talk area (dark blue area) and whose wavelengths are apart (distant points from the diagonal line of the figure). Triple wavelength measurement is expected to have a good noise performance because it solves an overdetermined linear system of equations to obtain the hemoglobin changes. However, triple wavelength measurement has not been thoroughly evaluated. This remains for future work.

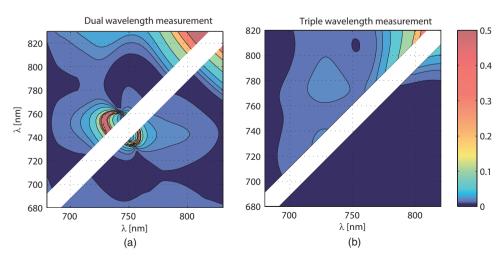


Fig. 6 Absolute values of Uludag's cross-talk measure of HbO to HbR, $C_{HbO\rightarrow HbR}$, by (a) dual and (b) triple wavelength measurement. They show the averaged cross-talk values when skull thickness values of 4, 7, and 10 mm were assumed.

Acknowledgment

The authors thank the Photon Migration Imaging Laboratory at Massachusetts General Hospital for the Monte Carlo simulation code (available at http://www.nmr.mgh.harvard.edu/ PMI/index.htm).

References

- 1. E. M. C. Hillman, "Optical brain imaging in vivo: techniques and applications from animal to man," *J. Biomed. Opt.* **12**(5), 051402 (2007).
- Y. Hoshi, "Functional near-infrared spectroscopy: current status and future prospects," J. Biomed. Opt. 12(6), 062106 (2007).
- 3. H. Obrig and A. Villringer, "Beyond the visible—imaging the human brain with light," *J. Cereb. Blood Flow Metab.* 23, 1–18 (2003).
- J. C. Hebden, "Advances in optical imaging of the newborn infant brain," *Psychophysiology* 40, 501–510 (2003).
- S. J. Matcher, C. E. Elwell, C. E. Cooper, M. Cope, and D. T. Delpy, "Performance comparison of several publishing tissue near-infrared spectroscopy algorithms," *Anal. Biochem.* 227, 54–68 (1995).
- K. Uludag, M. Kohl, J. Steinbrink, H. Obrig, and A. Villringer, "Cross-talk in the Lambert–Beer calculation for near-infrared wavelengths estimated by Monte Carlo Simulations," *J. Biomed. Opt.* 7(1), 51–59 (2002).
- 7. G. Strangman, M. A. Franceschini, and D. A. Boas, "Factors affect-

ing the accuracy of near-infrared spectroscopy concentration calculations for focal change in oxygenation parameters," *Neuroimage* **18**, 865–879 (2003).

- Y. Hoshi, M. Shimada, C. Sata, and Y. Iguchi, "Reevaluation of nearinfrared light propagation in the adult human head: implications for functional near-infrared spectroscopy," *J. Biomed. Opt.* 10(6), 064032 (2005).
- K. Uludag, J. Steinbrink, A. Villringer, and H. Obrig, "Separability and cross-talk: optimizing dual wavelength combinations for nearinfrared spectroscopy of the adult head," *Neuroimage* 22, 583–589 (2004).
- N. Okui and E. Okada, "Wavelength dependence of cross-talk in dual-wavelength measurement of oxy- and deoxy-hemoglobin," *J. Biomed. Opt.* **10**(1), 011015 (2005).
- M. Hiraoka, M. Firbank, M. Essenpresis, M. Cope, S. R. Arridge, P. van der Zee, and D. T. Delpy, "A Monte Carlo investigation of optical pathlength in inhomogeneous tissue and its application to nearinfrared spectroscopy," *Phys. Med. Biol.* 38, 1859–1876 (1993).
- Y. Fukui, Y. Ajichi, and E. Okada, "Monte Carlo prediction of nearinfrared light propagation in realistic adult and neonatal head models," *Appl. Opt.* 42, 2881–2887 (2003).
- 13. Y. Yamashita, A. Maki, and H. Koizumi, "Wavelength dependence of the precision of noninvasive optical measurement of oxy- and deoxy-, and total-hemoglobin concentration," *Med. Phys.* 28, 1108–1114 (2001).