

Integrated autofluorescence endoscopic imaging and point-wise spectroscopy for real-time *in vivo* tissue measurements

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Abstract. We report on the development of an integrated point-wise spectroscopy and autofluorescence (AF) endoscopic imaging technique for real-time *in vivo* tissue measurements at endoscopy. We implement a unique point spectrum optical design to realize real-time AF imaging and AF or diffuse reflectance (DR) spectroscopy measurements from a small area of tissue of interest on the AF image. We demonstrate that both the AF image and the point-wise AF/DR spectra can be simultaneously acquired from the oral cavity *in vivo* within 0.1 s, suggesting the potential of the integrated spectroscopy and endoscopic imaging technique developed to facilitate *in vivo* tissue diagnosis and characterization at endoscopy. © 2010 Society of Photo-Optical Instrumentation Engineers. [DOI: 10.1117/1.3475955]

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Head and neck cancer is one of the most common malignancies worldwide due to its high incidence and high mortality rates. In the United States, more than 35,000 new cases of head and neck malignancies were reported in 2009, accounting for approximately 3% of all newly diagnosed cancers.¹ In Singapore, over 2000 patients between 35 and 50 years old have been diagnosed with head and neck cancer from 1998 to 2002.² Early diagnosis and localization of head and neck cancer with effective treatment is critical to decreasing the mortality rates. However, identification of early cancer can be difficult using conventional white-light reflectance (WLR) endoscopy, which relies heavily on visualization of tissue morphological changes associated with neoplastic transformation. Subtle tissue changes may not be apparent, limiting its diagnostic accuracy. Positive endoscopic biopsy is the standard means for head and neck cancer diagnosis, but it is invasive and impractical for screening high-risk patients who may have multiple suspicious lesions. Hence, it is highly desirable to develop advanced optical techniques to complement WLR en-

doscopy for improving early cancer diagnosis and characterization during clinical examinations.

In the past two decades, the autofluorescence (AF) imaging technique, which is capable of detecting changes in the endogenous fluorophores and morphological architectures of tissue, has been developed to significantly improve the diagnostic sensitivity of early neoplastic lesions at endoscopy, but AF imaging still suffers from moderate diagnostic specificities.³ Optical spectroscopic techniques, such as AF spectroscopy and diffuse reflectance (DR) spectroscopy, which provide information about tissue optical properties (e.g., absorption and scattering coefficients), morphologic structures, endogenous fluorophore distribution, blood content (e.g., hemoglobin), and oxygenation associated with neoplastic transformation, have been comprehensively investigated for *in vitro* or *in vivo* precancer and cancer diagnosis in various organs with high diagnostic specificity.⁴⁻⁷ The combination of AF imaging with optical spectroscopy offers a potential of providing both high diagnostic sensitivity and specificity for cancer tissue diagnosis and detection.⁶⁻⁹ In this letter, we report on the development of an integrated point-wise spectroscopy (AF/DR) and AF endoscopic imaging technique for real-time *in vivo* tissue measurements at endoscopy. Different from previous work that combined imaging with spectroscopy limited to spectral measurements at the centroid of the endoscopic field of view,^{8,9} in our configuration, the *in vivo* point-wise AF/DR spectra can be quickly acquired from any specific areas of the imaged tissue of interest under the AF/WLR imaging guidance during endoscopic examination.

Figure 1 shows the schematic of the integrated point-wise spectroscopy and AF endoscopic imaging technique developed for *in vivo* tissue measurements at endoscopy. This system consists mainly of a dedicated 300-W xenon short arc lamp coupled with two customized bandpass (BP) filters (BP1: 375–440 nm for AF excitation; BP2: 400–700 nm for WL illumination) for AF/DR spectroscopy and imaging; a medical endoscope (Hopkins II 7230BP, Karl Storz, Germany); a sensitive three-chip charge-coupled device (CCD) camera [red (R) channel (600–700 nm); green (G) channel (500–580 nm); and blue (B) channel (400–480 nm); 752 × 582 pixels; Tricam SL II, Karl Storz, Germany]; a spectrograph equipped with a CCD detector (FWHM of ~1.5 nm with a 600 gr/mm holographic grating; USB2000, Ocean Optics, Inc., Dunedin, FL); and a specially designed point spectrum optical adapter (inset in Fig. 1) for realizing simultaneous *in vivo* endoscopic imaging and point-wise AF/DR spectral measurements on the specific areas of the imaged tissue of interest. The customized optical adaptor comprises three lenses ($f=50$ mm); a thin quartz glass plate ($30 \times 30 \times 1$ mm³) coated with a gold mirror (diameter of 100 μ m; reflection of ~99% in 400–1000 nm); and a 3-D motorized translational stage (travel range: 13 mm; 8MT184-13, Standa, Inc., Lithuania) for controlling the rapid movement of the gold mirror to realize the spectral measurements on the points of interest of the imaged tissue. For simultaneous AF imaging and spectroscopy measurements, the filtered blue excitation light (375–440 nm) is conducted into the endoscope via a flexible fiber optic light guide and shines onto the tissue with an incident power of 35 mW on the fiber tip of the endo-

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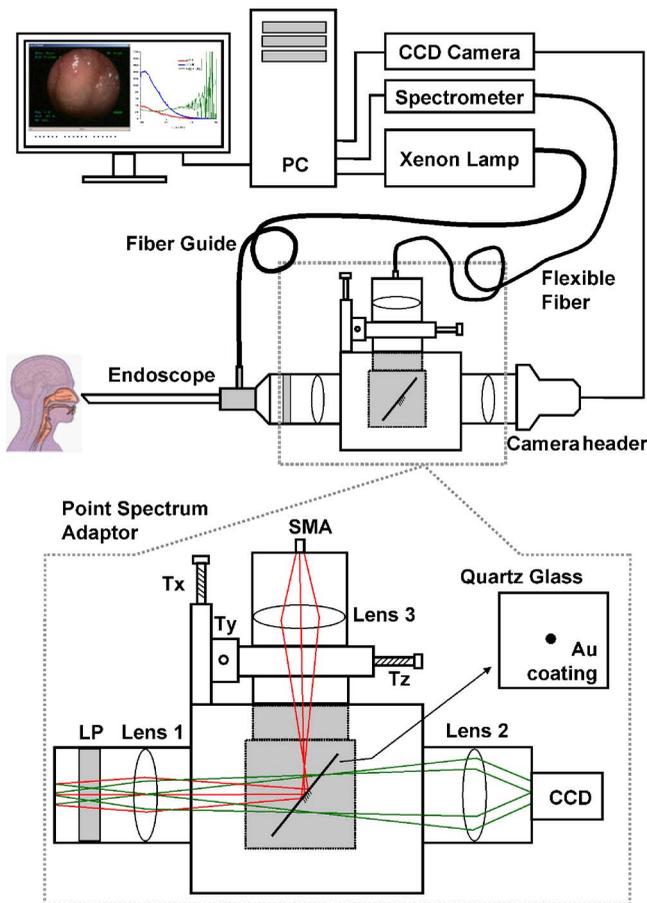
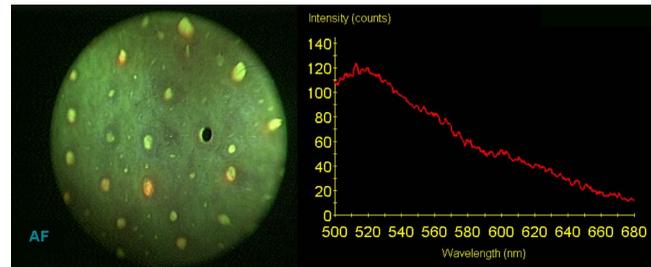


Fig. 1 Schematic of the integrated point-wise spectroscopy and autofluorescence (AF) imaging system for *in vivo* tissue measurements at endoscopy. Tx, Ty, and Tz stand for translational stages.

scope. AF emitted from the tissue is collected by the same fiber tip of the endoscope, then is coupled into the customized optical adaptor by passing through a long-pass (LP) filter (cut off at 480 nm) for removing the interference of the excitation light scattered from tissue, and then is focused onto the quartz glass plate, which is positioned at the interim imaging plane of lens 1 with an orientation of 45 deg with respect to the incident light direction. The tissue fluorescence light passes through the 45-deg oriented glass plate and is focused onto the three-chip CCD camera through lens 2 for fluorescence imaging measurements. Meanwhile, a very small portion of tissue fluorescence is reflected from the 100- μm gold mirror coated on the quartz plate and focused onto a 100- μm fiber via lens 3, which is connected to the spectrograph for fluorescence spectroscopic measurements. Further, an automatic motorization of the small gold mirror coated on the quartz plate together with the point-wise spectral measurement module enables a rapid movement of the dark spot (of ~ 0.5 mm in diameter due to the reflection of gold mirror in the point spectrum optical adaptor) on the image to any spot of the imaged tissue of interest (see Video 1). Hence, the AF imaging and point-wise AF spectroscopy can be simultaneously acquired from the same imaged tissue without introducing an optical fiber catheter into the instrument channel of an endoscope, as in conventional endoscopic spectral measurements, which prolong the endoscopic operation procedures. Similarly, the



Video 1 Video illustrating simultaneous AF imaging and point-wise AF spectral measurements of the cheek *in vivo* in real-time during AF endoscopic imaging (QuickTime, 11 MB). [URL: <http://dx.doi.org/10.1117/1.3475955.1>].

simultaneous WLR imaging and point-wise DR spectroscopy on the same tissue can also be realized simply by switching the excitation light filter to the white-light illumination mode (BP2: 400–700 nm) and removing the 480-nm LP filter in the customized optical adaptor. In this work, we have also developed a LabView-based software for controlling real-time endoscopic image (WLR/AF) acquisition and point-wise spectral measurements and processing (e.g., wavelength calibration, system spectral response calibration, CCD dark-noise subtraction, signal saturation detection, etc.). Both the live AF/WLR image and the AF/DR spectrum can be simultaneously displayed on the computer monitor for real-time review and stored in the computer for further analysis.

We have applied the integrated point-wise spectroscopy and endoscopic imaging technique developed for *in vivo* tissue measurements in the head and neck. Figure 2(a) shows an example of *in vivo* WLR images and the corresponding DR spectra of different tissue sites (i.e., chin, buccal mucosa, dorsal of the tongue, and lower lip) simultaneously acquired from a healthy volunteer under the white-light illumination mode. Point-wise DR spectra from different anatomical locations [dark spots in the WLR images in Fig. 2(a)] in the oral cavity can be acquired within 10 ms, and the absorption peaks (e.g., 420, 540, and 580 nm) attributed to hemoglobin absorptions in the vessels can be clearly identified, but with large absorption variations among different tissue locations.

Swapping the excitation filter to the blue BP filter (375–440 nm) in the xenon lamp for tissue fluorescence excitation, *in vivo* tissue AF images and point-wise AF spectra can also be simultaneously acquired from the head and neck. Figure 2(b) shows the representative *in vivo* AF images and AF spectra of different locations in the oral cavity from a healthy volunteer. Obviously, AF images [Fig. 2(b)] that contain information about endogenous fluorophore distributions in tissue provide a higher image contrast as compared to WLR images [Fig. 2(a)]. High-quality *in vivo* tissue AF spectra can be acquired within 0.1 s from the dark spot areas on the AF images [Fig. 2(b)]. Again, AF spectra from different anatomical tissue locations also vary, revealing the differences in concentrations of endogenous fluorophores among different tissue locations. For instance, the prominent fluorescence peak at 535 nm for flavins is observed in all different tissues, but a much stronger fluorescence at 630 nm for protoporphyrins is found, particularly in the chin and the tongue.

By rapidly moving the gold reflection mirror in the point spectrum optical adaptor, we also demonstrate the ability of

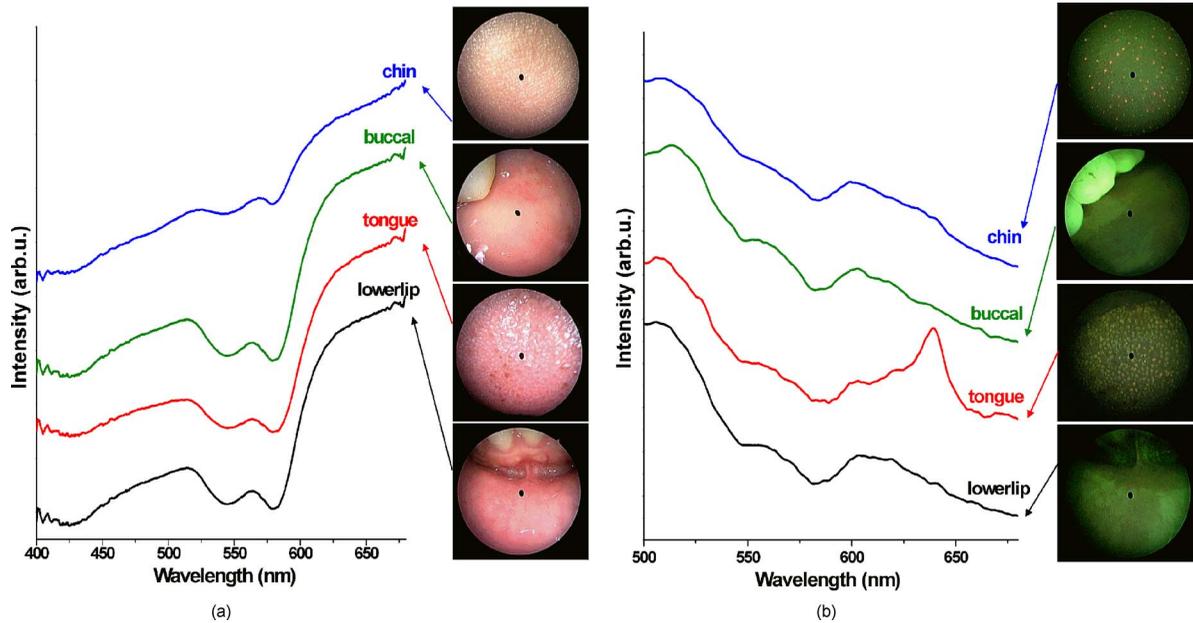


Fig. 2 (a) *In vivo* white-light images and the corresponding diffuse reflectance (DR) spectra from different anatomical locations (chin, buccal mucosa, dorsal of the tongue, and lower lip) simultaneously acquired from a healthy volunteer. (b) Comparison of *in vivo* AF images and the corresponding point-wise AF spectra from different anatomical locations (chin, buccal mucosa, dorsal of the tongue, and lower lip) simultaneously acquired from a healthy volunteer. Note that each DR spectrum is acquired within 10 ms, whereas the AF spectrum is acquired within 0.1 s.

the integrated endoscopic imaging and spectroscopy technique developed for pinpointing the spectral properties of specific areas of interest on the imaged tissue. [Video 1](#) illustrates the *in vivo* AF image of the cheek acquired together with simultaneous AF spectral measurements on different spots of the tissue imaged in real-time during rapid scanning of the gold reflection mirror (shown as a dark spot in the AF image). The point-wise spectral measurements across the entire image size of ~ 10 mm can be quickly completed within 2 to 3 s, making the *in vivo* AF measurements feasible in clinical settings. *In vivo* AF spectral differences of different spots on the same imaged tissue can also be clearly identified ([Video 1](#)), indicating the capability of our developed technique for revealing the inhomogeneity of endogenous fluorophore distributions in tissue. Our point spectrum adapter design associated with rapid scanning and optics allows for a more convenient spectral measurement on the point of interest of the entire imaged tissue for spectral analysis and may have a significant impact on practical clinical applications.

In conclusion, an integrated autofluorescence endoscopic imaging and point-wise spectroscopy system has been built and evaluated in the head and neck. The simultaneous acquisition of an endoscopic AF image and an AF/DR spectrum from a specific area of imaged tissue *in vivo* can be realized within 0.1 s, which may facilitate rapid, noninvasive, *in vivo* tissue diagnosis and characterization in clinical settings. One notes that the unique AF image-guided point-wise spectroscopy technique developed in this work can also be readily adapted to study other internal organs *in vivo* by using different flexible medical endoscopes (e.g., bronchoscope, colonoscope, gastroscope, etc.).

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